Acute Myelofibrosis and Malignant Hypercalcemia

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A 33 year old man presented with symptoms of one week's duration; he had a serum calcium of 22.5 mg/dl and a markedly hypercellular bone marrow. Despite therapy with saline diuresis, furosemide, mithramycin, total parathyroidectomy and corticosteroids, symptomatic hypercalcemia was poorly controlled. Inappropriate serum parathyroid hormone (PTH) levels were found before and after parathyroidectomy whereas assays of the peripheral blood for osteoclast-activating factor and prostaglandin E (PGE2) were negative. An elevated leukocyte alkaline phosphate level, the inability to aspirate marrow, the marked generalized hyperplasia of all hematopoietic marrow elements, the focal accumulations of blastic cells and increasing reticulin fiber formation led to the diagnosis of acute myelofibrosis. A single course of cytosine arabinoside and thio-guanine therapy was followed by profound hyperphosphatemia, hypocalcemia and death. The rarity of hypercalcemia with myeloproliferative disorders is documented by a review of the world literature, and the possible mechanism for hypercalcemia in this patient is discussed.

Our purpose here is to describe a young man who presented with severe hypercalcemia as the first manifestation of an unusually acute and uncommon myeloproliferative disorder.

CASE REPORT

A 33 year old Puerto Rican man was in excellent health until 10 days before his admission when he began to experience fatigue and dull, nonradiating bilateral chest and upper abdominal pain; the pain was gradual in onset and was followed some days later by nausea, vomiting, polyuria, polydipsia and nocturia. He denied use of medications or exposure to toxic substances, and his past and family history were noncontributory.

On admission he appeared acutely ill and lethargic, and he was nauseated and groaning with discomfort. Vital signs were normal and the physical examination disclosed no abnormalities except for diffuse abdominal tenderness. Lymph nodes, liver and spleen were not palpable.

Laboratory data on admission included urinalysis, complete blood count, platelet count, serum sodium, potassium, carbon dioxide, blood urea nitrogen, glucose and amylase levels. X-ray films of the chest and abdomen were within normal limits.

By the next morning the patient was semicomatose, areflexic and without bowel sounds. The blood calcium and phosphate samples drawn the previous day demonstrated respective values of 14.7 and 4.3 mg/dl and when redrawn revealed levels of 22.5 and 3.8 mg/dl. A repeat serum amylase was again normal. An initial trial of rapid saline infusion resulted in no significant decrease of the calcium level and intravenously administered furosemide, up
to 80 mg/hour, was added. Despite the marked increase in urinary excretion of calcium (2,332 mg in 7.35 liters over a 12 hour period), the serum calcium level failed to fall below 17 mg/dl and the blood urea nitrogen and creatinine levels began to rise (Figure 1). The administration of mithramycin as a single intravenous dose of 50 μg/kg was followed by a fall in the serum calcium level to 7.9 mg/dl 48 hours later. Clinical improvement was rapid; the patient became alert and essentially asymptomatic, and physical findings became normal. The blood urea nitrogen returned to normal, but the creatinine level remained mildly elevated for the balance of his illness.

Serum and urine protein electrophoresis, roentgenographic studies including a skeletal survey, a series of the upper and lower gastrointestinal tract, oral cholecystogram and intravenous pyelography, and repeated complete blood counts, electrolytes and urinalysis were within normal limits. An elevation in the serum amylase level was noted which persisted during the remainder of hospitalization with fluctuations between 226 and 1,722 IU (normal of 60 to 250 IU). Urine amylase levels were proportionately elevated and the amylase clearance was normal [1]. A sternal marrow aspirate was obtained without difficulty. Both clot sections and smears demonstrated a moderately hypercellular bone marrow with moderate myeloid hyperplasia and scattered ill-defined foci of small immature round cells with scanty cytoplasm, round or somewhat ovoid nuclei and often 1 or 2 nucleoli, approximating the appearance of hematopoietic stem cells as described by Dicke et al. [2]. On a differential count of the smears, these cells accounted for 22 per cent of all cells with another 17 per cent identified as bone marrow "lymphocytes." A silver stain was negative for reticulin. The complete blood count and peripheral blood cell morphology were normal with a leukocyte alkaline phosphatase score of 165 IU (normal 13 to 130 IU). Five days later an easily obtained bone marrow aspirate and Jamshidi trephine biopsy specimen from the right posterior iliac crest demonstrated a markedly hypercellular marrow with virtually no fat, generalized hyperplasia of erythroid, myeloid and megakaryocytic cell lines without evidence of maturation arrest or the immature elements observed in the sternal specimen. The silver stain of the biopsy specimen revealed some focal areas of 1+ to 2+ reticulin staining immediately adjacent to bony trabeculae but otherwise was negative. Unfortunately, the marrow culture preparation produced no usable mitoses for karyotypic analysis. Except for moderate anemia, the complete blood count and peripheral morphology remained within normal limits.

On the eighth hospital day, the serum calcium level began to rise (Figure 1) despite the continuous vigorous saline diuresis. The patient again became lethargic and experienced generalized muscular pain. A bone scan with technetium 99m pertechnate disclosed no abnormalities except for two focal areas of increased uptake in the rib cage which subsequently proved to be old healed rib fractures. In the face of no definitive cause for the hypercalcemia, the patient underwent surgical exploration of the neck on the 17th day and four parathyroid glands, normal in size and histology, were identified in the specimen submitted for tissue examination. Despite the total parathyroidectomy and continuing massive diuresis, the calcium level continued to rise postoperatively, necessitating repeated mithramycin therapy. On the 14th day a biopsy specimen of a bleeding rectal ulceration was obtained which showed extensive underlying soft tissue calcium deposits. Coagulation studies, including platelet function studies, disclosed no abnormalities. On the 16th day a repeat bone marrow aspiration was attempted from several different segments of the sternum, but only a few drops of sinusoidal blood could be aspirated from any one site. The differential count of this scanty specimen was identical to that in the first sternal aspirate. A tiny specimen obtained by sternal trephine suggested myelofibrosis and demonstrated increased reticulin formation. Except for a few myelocytes and metamyelocytes, and rare nucleated red cells, morphologically the peripheral blood remained normal with no evidence of tear drop forms. Serum levels of muramidase, folate and vitamin B12 were within normal limits.

Four units of packed red blood cells were given at the time of the parathyroidectomy and in the immediate postoperative period, but the hemoglobin level continued to fall at a rate greater than expected from the degree of intravenous blood loss, suggesting some combination of decreased red cell production and accelerated destruction, however, insufficient to depress the haptoglobin or increase the serum bilirubin. The Coombs' antiglobulin test result was negative, and the maximum recorded reticulocyte count was 1.6 per cent. The patient subsequently received 4 more units of packed red cells during his illness.

PTH levels, as determined by radioimmunoassay by the Mayo Medical Laboratories from blood samples drawn before
and five days after total parathyroidectomy, were inappropriately high with values of 25 and 28 μl eq/ml, respectively, when serum calcium levels were 14 to 15 mg/dl. Ordinarily, no PTH should be detectable under these conditions [3]. When the diagnosis of an unusual and rapidly progressive acute myeloproliferative disorder became apparent, the administration of prednisone, 169 mg, and allopurinol, 300 mg, daily was begun on the 17th hospital day without any effect on the calcium level; on the 19th day a single dose of cytosine arabinoside, 3 mg/kg intravenously, and thioguanine, 2.5 mg/kg orally, was given. Early on the 20th day another dose of mitomycin C was given intravenously. Later that morning the patient became extremely tachypneic with evidence of pulmonary edema followed by cardiac arrest and complete atrioventricular dissociation. He was successfully resuscitated but subsequently remained anuric; severe hyperphosphatemia of 17 mg/dl and hypocalcemia of 6.7 mg/dl developed, and he died early on the 21st hospital day, just one month after the initial onset of symptoms. A peripheral blood sample obtained shortly before his death was assayed for osteoclast activating factor (OAF) by Dr. Lawrence G. Raisz, utilizing the fetal rat in vitro tissue culture system [4], and was found negative. In addition, prostaglandin E2 was not detectable in the peripheral plasma.

Gross postmortem examination revealed marked pulmonary edema and bilateral pleural effusions but was otherwise unremarkable. A thorough search failed to reveal any additional parathyroid tissue. The spleen was slightly enlarged with passive congestion, and the lymph nodes and liver appeared normal. The skeleton was grossly normal except for two old healed rib fractures with considerable callous formation. Postmortem bone marrow aspiration was attempted from at least a dozen areas and produced dry taps except from two sites. On Wright's-stained smears, one of these demonstrated hypercellular normal appearing bone marrow elements with large numbers of osteoclasts. Occasional frankly megaloblastic erythroid precursors were seen and were thought to be related to the cytosine arabinoside therapy. The other site revealed numerous osteoclasts scattered among a monotonous field of small undifferentiated blast cells with scanty cytoplasm similar to those seen previously during life in the sternum (Figure 2). On section the bone marrow was markedly hypercellular. In some areas increases in all hematopoietic cell lines were seen with a modest or no shift to the left, whereas other areas were replaced by small mononuclear blast cells. The bony trabeculae demonstrated extensive erosion and scalloping (Howship's lacunae) with large numbers of adjacent osteoclasts (Figure 3). Moderate to marked reticulin formation was demonstrated throughout the entire bone marrow although collagen fibers were not seen. Microscopically the lungs revealed marked pulmonary edema with focal areas of bronchopneumonia. Extensive deposition of calcium was demonstrated in the myocardium, stomach, intestines, kidneys, bladder, prostate and skeletal muscle with a marked predilection for muscle, whether smooth, cardiac or striated. The pancreas was normal. Except for some small foci of myeloid precursors with a few normoblasts without megakaryocytes in the red pulp of the spleen, there was no evidence of extramedullary hematopoiesis or neoplastic infiltration outside the bone marrow.

COMMENTS

Although hypercalcemia is a common complication of certain malignancies [5–7], with the exception of multiple myeloma, the association with myeloma or lymphoproliferative disorders is infrequent [8,9]. Osteolytic lesions secondary to hematopoietic or lymphoid neoplasia are quite uncommon other than in multiple myeloma and in some cases of acute lymphoblastic leukemia in children, Hodgkin's disease, large cell ("histiocytic") and Burkitt's lymphomas, despite the fact that the marrow frequently is extensively infiltrated by neoplastic tissue [10,11]. Thus, osteolytic activity and hypercalcemia are not simply related to intramedullary
### Table I
Reported Cases of Hypercalcemia Associated with Myeloproliferative Disorders

<table>
<thead>
<tr>
<th>Case No.</th>
<th>Reference</th>
<th>Date</th>
<th>Age (yr)</th>
<th>Peak Calcium (mg/dl)</th>
<th>Disorders</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Doub, Hartman [38]</td>
<td>1935</td>
<td>51, F</td>
<td>13.7</td>
<td>&quot;Ateumatic myelosis&quot; (myelofibrosis?)</td>
</tr>
<tr>
<td>2</td>
<td>Doub, Hartman [38]</td>
<td>1935</td>
<td>17, M</td>
<td>11.5</td>
<td>Acute granulocytic leukemia</td>
</tr>
<tr>
<td>3</td>
<td>Kronfield, Reynolds [41]</td>
<td>1964</td>
<td>14, M</td>
<td>18.7</td>
<td>Acute granulocytic leukemia</td>
</tr>
<tr>
<td>4</td>
<td>Jordan [42]</td>
<td>1966</td>
<td>32, M</td>
<td>13.1</td>
<td>Acute granulocytic (or undifferentiated) leukemia</td>
</tr>
<tr>
<td>5</td>
<td>Jordan [42]</td>
<td>1966</td>
<td>47, M</td>
<td>11.9</td>
<td>Acute granulocytic leukemia</td>
</tr>
<tr>
<td>7</td>
<td>Ballard, Marcus [43]</td>
<td>1970</td>
<td>40, M</td>
<td>21.6</td>
<td>Chronic granulocytic leukemia</td>
</tr>
<tr>
<td>8</td>
<td>Steinberg et al. [44]</td>
<td>1971</td>
<td>40, M</td>
<td>16.0</td>
<td>Chronic granulocytic leukemia in acute blastic phase</td>
</tr>
<tr>
<td>9</td>
<td>Haskell et al. [45]</td>
<td>1971</td>
<td>44, F</td>
<td>12.1</td>
<td>Chronic granulocytic leukemia in acute blastic phase</td>
</tr>
<tr>
<td>10</td>
<td>Haskell et al. [45]</td>
<td>1971</td>
<td>52, M</td>
<td>13.8</td>
<td>Chronic granulocytic leukemia in acute blastic phase</td>
</tr>
<tr>
<td>11</td>
<td>Neiman, Li [12]</td>
<td>1972</td>
<td>13, M</td>
<td>17.0</td>
<td>Acute undifferentiated leukemia</td>
</tr>
<tr>
<td>12</td>
<td>Palva, Salokannel [46]</td>
<td>1972</td>
<td>15, M</td>
<td>15.6</td>
<td>Acute granulocytic leukemia</td>
</tr>
<tr>
<td>13</td>
<td>Allen et al. [47]</td>
<td>1973</td>
<td>80, F</td>
<td>14.2</td>
<td>Acute myelomonocytic leukemia</td>
</tr>
<tr>
<td>14</td>
<td>Kawasaki et al. [48]</td>
<td>1973</td>
<td>24, F</td>
<td>18.0</td>
<td>Acute granulocytic leukemia</td>
</tr>
<tr>
<td>15</td>
<td>Licht et al. [39]</td>
<td>1973</td>
<td>40, F</td>
<td>14.2</td>
<td>Chronic granulocytic leukemia with terminal myelofibrosis</td>
</tr>
<tr>
<td>16</td>
<td>Souillet et al. [49]</td>
<td>1973</td>
<td>14, F</td>
<td>16.5</td>
<td>Acute granulocytic leukemia</td>
</tr>
<tr>
<td>17</td>
<td>Greul, Lozano-Tonkin [50]</td>
<td>1974</td>
<td>63, M</td>
<td>16.6</td>
<td>Acute aleukemic monocytic leukemia</td>
</tr>
<tr>
<td>18</td>
<td>McKeen [51]</td>
<td>1974</td>
<td>49, M</td>
<td>14.6</td>
<td>Acute granulocytic leukemia</td>
</tr>
<tr>
<td>19</td>
<td>McKeen [51]</td>
<td>1974</td>
<td>58, M</td>
<td>15.6</td>
<td>Chronic granulocytic leukemia in acute blastic phase</td>
</tr>
<tr>
<td>20</td>
<td>Present case</td>
<td>1974</td>
<td>33, M</td>
<td>22.5</td>
<td>Acute myelofibrosis</td>
</tr>
</tbody>
</table>

Cell numbers and mechanical erosion of bone, but rather to cell type and presumably one or more cell products which lead to these sequelae. Factors suggested as causes of osteolysis and hypercalcemia in malignancy are PTH or PTH-like substances [3,12], prostaglandin E2 [13], a specific osteoclastic activating factor [14,15], some other non-PTH hormone [16], or an imbalance of the various normal factors operating on skeletal and mineral homeostasis [17,18]. The inappropriately high serum PTH level before and after total parathyroidectomy in our case suggests that PTH or a PTH-like substance was produced by the abnormal hematopoietic cells. The negative assay of the peripheral blood for osteoclastic activating factor activity does not rule out the strictly intramedullary production of such a factor or its inactivation or destruction in the peripheral blood. The in vitro demonstration by Mundy et al. [16] of an osteoclastic activating factor from myelomatous tissue was not accompanied by any such substance detectable in the peripheral blood of the same patients. However, to date, all cells reportedly producing osteoclastic activating factor have been lymphoid in origin [19,20]. Whether some inducer of prostaglandin E2 synthesis also was present cannot be absolutely ruled out since no trial with indomethacin, a potent inhibitor of prostaglandin E synthesis, was attempted in this patient. However, the prostaglandin E2 level in the peripheral blood was not elevated.

Acute myelofibrosis is a rare and somewhat controversial myeloproliferative disorder only recently delineated as an entity by Lewis and Szm [21] and others [22-31], although similar cases can be found in the older literature [32-35]. Some of these patients demonstrated an accelerated development of myelofibrosis otherwise morphologically or clinically indistinguishable from the classic more insidious variety. The other more distinctive group displayed coexistent features of myelofibrosis and a prominent blastic component, at least in focal areas, distinct from the 6 to 20 percent of the patients in whom an acute blastic terminal phase develops only years after the onset of classic myelofibrosis with myeloid metaplasia [36] or from those patients with acute leukemia and increased marrow reticulin fiber formation [37]. Our patient's bone marrow picture of a striking pancytosis with the generalized reticulin fiber formation, focal accumulations of blast cells and elevated leukocyte alkaline phosphatase level is consistent with the diagnosis of acute myelofibrosis. As noted by others [27], the rapidity of the course and early death of the patient probably accounts for the lack of significant extramedullary hematopoiesis or the development of collagen fibers. In the absence of symptomatic hypercalcemia, clinical and laboratory abnormalities revealing the nature of the underlying myeloproliferative disorder might not have become apparent for some time.
Hypercalcemia has not been reported as a complication of myelofibrosis except possibly for a case described by Doub and Hartman [38] in 1935 as “aleu-
lemic myelosis,” one of the older designations for myelofibrosis with myeloid metaplasia, although apparently no special stains for reticulin fibers or collagen were made. The other possible exception is a patient with chronic granulocytic leukemia and terminal myelofibrosis [39]. Unfortunately, an autopsy was denied in that patient, and a terminal nonleukemic blastic transformation as found in most of the patients evaluated by Grahnick et al. [40] could not be ruled out. The incidence of hypercalcemia with all myeloproliferative disorders is extremely low. A review of the world literature uncovered only 19 cases reported to date in addition to the present case (Table I). The mechanism leading to hypercalcemia is not indicated in any of the 19 except for the case of acute leukemia described by Neiman and Li [12] in which excessive PTH or PTH-like activity was demonstrated. In at least 17 of the 20 cases the presence of a sizable population of blastic cells was documented when the hypercalcemia developed. This suggests that the potential to produce substances leading to osteolysis tends to be a property of the earlier cellular stages of hematopoiesis. This is supported by the extreme rarity of frankly osteolytic lesions proved microscopically to be due to chronic myeloproliferative disorders lacking a significant blastic component [52–54].

In about half of the cases of classic “agogenic” myeloid metaplasia, not only myelofibrosis but also osteosclerosis is demonstrated [55–57]. Contrary to the original concept of the myeloproliferative syn-
dromes [58], the fibroblastic response in myelofibrosis appears not to be neoplastic but rather a secondary reactive phenomenon [26], similar to the desmoplastic reaction seen with many other neoplasms [59–63]. The proliferating clone of hematopoietic cells in myelofi-
brosis and in some cases of chronic granulocytic leukem-
ia [40] and polycythemia vera [64] stimulate local fibroblastic and less often osteoblastic activity by as yet un-
defined mechanisms. It is unknown whether these responses are mediated through cell-cell interaction, elaboration of toxic substances or short range hor-
mones. Many of the hormonal substances produced by hematopoietic or lymphoid cells, such as osteoclastic activating factor [14,15,19,20] and certain other of the lymphokines [65], and the “chalone” involved in in-
tramural control of granulopoiesis [66] exert a strictly local effect due to either inactivation or dilution away from the site of production, making their isolation and identification difficult. According to current hyp-
thesis [17], osteoblasts derive from osteoclasts by a process “modulated” by mechanical stress, calcii-
tonin, inorganic phosphate, growth hormone, estrone and inhibited by PTH and osteoclastic activating factor. The enhanced proliferation of the progenitor of the osteoclasticosteoblastic-osteolytic cell line in mye-
lofibrosis and the simultaneous production of a PTH-like substance in our patient may account for the intensity of the osteoclastic and osteolytic response.

The terminal severe hypocalcemia in our patient could be accounted for by the inhibition of PTH-like action on the osteoclasts and osteocytes by mithra-
mycin, halting calcium mobilization [67], the cessation of further PTH production and the possible binding of plasma calcium by phosphates released from the cy-
toplasm of damaged or killed neoplastic cells. Marked hyperphosphatemia following chemotherapy has been well documented recently in cases of acute leukemia [68,69], lymphocytic lymphoma [70] and Burkitt’s lymphoma [71] and was considered to be the responsible mechanism in this case [72]. To what degree the terminal hyperphosphatemia contributed to the extensive deposition of calcium salts in the soft tissue found at autopsy is uncertain, in part because microscopic soft tissue calcification on biopsy was already evident earlier in the course, and in part because of the uncertainty of how hyperphosphatemia produces the fall in serum calcium levels [73].

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